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cleaning materials, solvents, paints, varnishes, Sterno fuel, formaldehyde solutions, antifreeze, gascoal, moonlight, wind washer liquid (30-40% methanol) and duplicate liquids depressant of the central nervous system, Methanol is potentially toxic in quantities as small as one mouth When metabolized liver alcohol and aldehyde dehydrogenase, methanol forms formaldehyde and formic acid, both of which are toxic eyes, CNS, and gastrointestinal affected formic acid is the main toxin that makes up much of the anionic rupture, metabolic acidosis, and the gastrointestinal tract. Lactic acid also promotes anion rupture Ofmic acid suppresses cytochrome oxidase in the fondue of the eye; Swelling of the axons in the visual disk and swelling lead to visual impairment formaldehyde has a short half-life, lasts only a few minutes Formal acid metabolizes much more slowly, and this bio-accumulates with significant vertigo of methanol intake, blurred vision, fuzzy, foggy, or blizzard-like visual impairment of respiratory depression/death. Auxiliary measures and provision of airway protection. Dialysis may be required to remove methanol and its main toxic metabolite, formate. (Although forced diuresis can be considered, as methanol is excreted renail - dialysis works better and has a lower risk of pulmonary edema, brain swelling, or acute respiratory distress syndrome). Attempting to correct acidosis using sodium bicarbonate is indicated if the pH is less than 7.20: Please note that patients may require large amounts. Alkaline pH makes it more likely that famic acid will exist as its anion (formate), which cannot access the central nervous system and the optic nerve as easily. Administering folic acid (leukovorin) 50 mg IV every 4 hours for a few days to the potentate folic acid depends on the metabolism of form acid to carbon dioxide and water. Consider ethanol infusion in any patient with unexplained osmolar rupture and/or elevated anion-gap metabolic acidosis, which is unaccounted for ethanol until a definitive diagnosis of denial of its introduction is made. delay in the treatment of methanol intoxication can lead to increased morbidity and mortality, and timely treatment is essential for full recovery. International Program on Chemical Safety Poison Information Monograph 335 Chemical methanol Aliphatic alcohol carbinol colonial spirit Colombian alcohol methanol methanol (Italian) methylated alcohol (s) methyl alcohol methyl hydroxide monohydroximetan pyroxylic wood wood wood napl alcohol alcohol alcohol. 1.4.1 CAS Number 67-56-1 1.4.2 Other NIOSH numbers: PC 1400000 Local only. It doesn't matter. The main risks are heavy metabolic acidosis with an increase in the rupture of anion caused by the accumulation of mimic acid and in the late stages also lactic acid. Acidosis and metabolite pulp cause depression/toxicity of the central nervous system and visual impairment, which may be Full blindness is possible. Targeted organs: central nervous system and retina. Neurological: Early signs of mild intoxication and drowsiness. Delays appear after 8 - 36 hours: headache, dizziness, drowsiness, coma, and sometimes convulsions. Extended pupils, with a sluggish or absent mild reflex, occur in conscious patients. Visual impairment: Vision becomes blurred or blacked out, and may be enough to impair light perception or cause complete blindness. There is a disturbance of the pupil's response to light, and a shrinking field of vision, a cattle, and flashing lights. Visual impairment can be permanent. Visual disc hyperemia is common in the acute stage. Other: Abdominal pain is common; acute pancreatitis can occur. Severe recurrent metabolic acidosis with an increase in anion rupture is caused by the accumulation of formate and is associated with the severity of poisoning. Osmolal rupture also usually rises in the early stages before methanol metabolizes into formic acid. The amilas serum can be elevated. Hyperkalemia can occur associated with metabolic acidosis, and red blood cells medium body volume can be increased. If the definition of methanol cannot be met, the anion and osmol rupture should always be calculated (Jacobsen and McMartin, 1997). The definition of formate in the blood and urine enismatic method can also be performed, but rarely available in the acute phase. First aid measure Acute poisoning occurs mainly when ingested. All cases of ingestion must be sent to the hospital. If exposure occurred when inhaled or in contact with the skin, remove the patient from exposure, remove contaminated clothing, and wash the skin and eyes thoroughly. Management Principles: Consider emptying the stomach of gastric tinghing after the recent (1 hour) pumping of large quantities. Correct metabolic acidosis with sodium bicarbonate, regulating the rate of administration in accordance with repeated and frequent measurement of acid-base condition. Administering ethanol or phomepisol as an antidote to inhibiting the formation of toxic metabolites. Hemodialysis removes methanol and its metabolites, and helps in the correction of metabolic acidosis. If specific treatment is started early enough, it can prevent death and irreversible damage to vision. Methanol was originally produced as a result of destructive wood distillation, but is now commonly produced from hydrogen and carbon monoxide or carbon dioxide, as well as by hydrocarbon oxidation (Windholz, 1983). Formula: CH3OH Molecular Weight: 32.04 3.3.1 Color Color, Smell: Pulp, Colorless Liquid, with a slight alcoholic smell. 3.3.2 Condition/Form Normal state at room temperature: liquid. 3.3.3 Boiling Point Description: 65 degrees Celsius Melting Temperature: 97.8 degrees Celsius Flash point: 12 degrees Celsius (54 degrees Fahrenheit) Auto-preparation temperature: 470 degrees Celsius (air and l): d20/4: 0.7915 Relative vapour density (air and l): 1.11 pairs of pressure pressure 25 degrees Celsius: 125 mm Hg. (16.2 KP) at 20 degrees Celsius: 94 mm Hg. (vol.% in the air): 6.0 to 36.5 Dangers associated with steam, its dissipation and possible ignition: the liquid is flammable and at normal room temperatures develops vapors that form explosive mixtures in a wide range of concentrations. The risk of inhaling fumes depends on the concentration and duration of exposure (Parmeggiani, 1983). Possible chemical reactions: violent reaction with oxidizing substances such as CrO3, Pb (ClO4)2, HClO4, P2O3, (Lewis, 1996). Fire hazards: dangerous when exposed to heat, flames or oxidative agents (Aquilonius et al., 1978). Spontaneous heating: no (Sax et al., 1989). Danger of explosion: modest when exposed to flames (Sax et al., 1989). See also the possible chemical reactions, above. In some countries, dye is added to methanol to distinguish it from ethanol (e.g. in the United Kingdom, purple dye). 4.1.1 Use of industrial/commercial products designed to remove non-home; Industrial - Another removal product; Industrial fuel/ignition source; Industrial - Fuel; Liquid Industrial solvent; Maintenance of industrial vehicles; Industrial - antifreeze (vehicle); Industrial - Fuel supplement; Industrial - Screenwash; Industrial industrial operation - Denaturant; Industrial chemicals used in synthesis; Otherwise, the household/leisure material for drawing the product is not specified; Domestic - stripper paint; Internal fuel/ignition source Domestic - fuel liquid; Maintenance of domestic vehicles; Domestic - antifreeze; Vehicle; Domestic - Screenwash; Domestic Food/Food Pollutant Drink - Chemical Food Pollutant 4.1.2 Description of Industrial Solvent. Raw materials for the manufacture of formaldehyde and methyl esters of organic and inorganic acids. Antifreeze for radiators and air brakes. Ingredient of gasoline and diesel oil antifreeze. Liquids for washing the windshield. Liquid fuel for small engines used in hobby Fuel for picnic furnaces / solder torches (Windholz; Merck Index, 1983) Solvent for ink, dyes, resins and adhesive ingredients for the removal of paint and varnish (Parmeggiani, 1983). A denature for ethanol, which is not intended for human consumption, in concentrations of up to 5% or more (accidental use). Most poisonings occur when ingested. Industrial exposure to methanol vapors is also possible. Sporadic cases of methanol poisoning are usually caused by accidental or suicidal use of a product that may be available in chemical laboratories, and sometimes at home. Poisoning epidemics create very serious problems and are usually the result of the reception of falsified perfumes. Exposure can occur in works related to different uses of the product. Workers should not be exposed to harmful concentrations of steam. However, there is ample evidence from the photographic film industry that repeated exposure to air level exceeding the 200 ppm threshold does not cause significant discomfort or illness (Finkel et al., 1983). In any case, persons with any CNS disorder and/or reduced visual acuity should not be exposed to methanol (Parmeggiani, 1983). Note: Since methanol poisoning often occurs as accidents involving many people, the doctor often deals with medicine catastrophes, and has to change the approach to treatment respectively (Jacobsen and McMartin, 1986). Poisoning can occur as a result of ingestion, inhalation or percutaneous absorption (Dutkiewicz et al., 1980; Kahn and Blum, 1979). Finished absorbed. Finished absorbed. Finished absorbed. It's possible. It's possible. There are no data. Methanol is easily absorbed from the gastrointestinal tract and respiratory tract, as well as by percutaneous pathway (Dutkiewicz et al., 1980; Kahn and Blum, 1979). After absorption, methanol is widely distributed in the total water of the body with a distribution volume of 0.6 to 0.7 liters/kg (Jacobsen and McMartin, 1997). There is no protein binding (Jacobsen and McMartin, 1986). Undissociated pulp easily crosses the hem-brain barrier. Therefore, aggressive alkaline treatment is important in order to ensure that most formic acids are separated (Jacobsen and McMartin, 1997). It spreads poorly in fatty tissues. The elimination of methanol has zero order at a rate of 8.5 g/hour, i.e. about half of ethanol (Jacobsen et al., 1988). If the metabolism of methanol is blocked by ethanol or fomepisol, the removal of methanol occurs very slowly (about 50 hours) and occurs in pulmonary and renal excretion (Brent et al., 2001, Jacobsen et al., 1983). Most methanol is converted into formaldehyde, mainly in the liver, by dehydrogenase alcohol. Formaldehyde is converted into aldehyde dehydrogenase and other enzymes (Jacobsen and McMartin, 1997). In monkeys and rats, methanol is additionally metabolized into carbon dioxide using a folic-dependent single-carbon pathway pool (Vale and Meredith, 1981; Gosselin et al., 1984; Jacobsen and McMartin, 1986). Studies have shown that the rate of oxidation is regulated by hepatic concentrations of tetrahydrofolate (Eells et al., 1982). Concentrations of tetrahydrofolate in the livers of monkeys and humans are significantly lower than in the liver of rats (Johlin et al., 1986), which probably explains the slower metabolism of milling in primates (Jacobsen and McMartin, 1986). Metabolism and methanol toxicity are characterized by important differences between species; in non-primate laboratory animals, methanol itself, rather than its metabolites, is a major toxic agent (Roe, 1982; Gosselin et al., 1984). (See also sections 6.4 and 7.2.2.) Methanol oxidation also occurs in the kidneys (Winchester, 1983). The metabolic rate does not depend on plasma concentration, slowly, and is about one-seventh that of ethanol. Full oxidation and methanol may take several days. Since ethanol has an affinity for alcohol dehydrogenase, which is at least 20 times more than methanol, it predominantly serves as a substrate for this enzyme (Gossel and Bricker, 1984). The administration of ethanol (or phomepisol) reduces the rate of methanol oxidation and delays its clinical and biochemical effects. Thirty percent of the ingested dose is excreted without changes in the airways. The kidney emits less than 5% of immutable methanol. Formate is found in urine 4 to 10 days after one exposure (Baselt, 1982a). The release of urine for 70 mg/24 hours confirms the diagnosis of methanol intoxication (Bozza-Marrubini et al., 1987). This is comparable to the average concentration of 2-30 mg/L in non-exposure subjects (Baselt, 1982a; Schaller and Tribeg, 1985). The average ratio of urine concentration in the blood to methanol in humans is 1.30 (Baselt, 1982a). Methanol poisoning is characterized by metabolic acidosis and eye damage. The toxicity of methanol is caused by its metabolites, not methanol itself. The severity of toxicity correlates with the degree of metabolic acidosis, not with the concentration of methanol (Jacobsen and McMartin, 1986; 1997; Schwartz et al., 1981). The mechanism of eye damage is not fully understood (Jacobsen and McMartin, 1997). Experimental results show that eye damage is due to the toxicity of formate, regardless of acidosis. However, acidosis probably causes faster development of eye damage (Jacobsen and McMartin, 1986; Martin-Amat et al., 1977; Martin-Amat et al., 1978). Severe metabolic acidosis is typical for methanol poisoning. Initially, acidosis is caused by the cream acid itself. At a more advanced stage, it is also supported by milk acidosis, which may be caused by the inhibition of cytochrome oxidase (see section 7.2.3.3) due to tissue hypoxia (Jacobsen and McMartin, 1986). Concentrations of pulp acid in the blood are proportional to the widening gap in the anion (Sejersted et al., 1983). For this reason, serum bicarbonate levels and baseline excess values are reliable early indicators of the severity of methanol poisoning (Bozza-Marrubini et al., 1987). Pathological results in the autopsy of the liver, kidneys and heart show parenchymatous degeneration. The lungs show desquamation of epithelial, emphysema, swelling, congestion and pneumonia. The brain can show swelling, hyperemia, petehia and heart attack of confuseds, which in severe cases can be hemorrhagic (McLean et al., 1980; Aquilonius, etc., 1980). The eye shows degenerative changes in the retina and swelling of the visual disk, and there may be optic nerve atrophy. The epithelium of the cornea can show degenerative changes. There was also pancreatic necrosis (Bennet et al., 1952). 7.2.1 Human data 7.2.1.1 Adults (volunteers Clinical Case Data) Volunteers After taking 4 ml methanol, urinary minoan acid levels reach a maximum of about 56 mg/l for 2 hours and then decline rapidly (Kendal and Ramanathan, 1953). The peak concentration of methanol in the blood of 117 mg/L (3.6 mmol/L) was reached 1 hour after an adult male volunteer weighing 78.5 kg swallowed 7 ml methanol (Leaf and zatan, 1952). Data on occupational effects in workers exposed to methanol at vapour concentrations of 85 to 134 ppm increased from an average of 13 mg/l in the morning to 20 mg/l by evening (Baselt, 1982a). Clinical data on cases of acute intake of 4 to 10 ml of methanol can cause permanent blindness (Vale s Meredith, 1981; Bozza-Marrubini et al., 1987; Gossel and Bricker, 1984; Litovic, 1986). However, individual susceptibility varies widely, possibly due to frequent simultaneous intake of ethanol and recovery after taking 500 to 600 ml was recorded (Gossel and Bricker, 1984; Litowitz, 1986). A retrospective analysis of data on large-scale methanol poisoning by falsified wine in Italy 1986 showed that no one with urine less than 200 mg/L developed any true symptoms or objective clinical signs. Other Human Data (Sax et al., 1989): Human inhaling TClO: 86,000 mg/m3: IRR Human Eye: 5 ppm. 7.2.1.2 A very limited experience of children with infants suggests that the clinical picture is similar in adults in all important ways (Gosselin et al., 1984), but that children may be more sensitive to toxic effects (Bozza-Marri et al., 1987). Poisoning occurred in a child at the age of 10 weeks, when methanol was mistaken for distilled water and mixed in the child's food (Wenzl et al., 1968). Child poisoning can occur as a result of skin absorption or inhalation (Litovitz, 1986). An 8-month-old baby died of methanol toxicity after a cold, rubbing his chest with olive oil and then applying warm methanol compresses (Kahn and Blum, 1979). 7.2.2 Appropriate oral rat animal data: LD50: 5628-13000 mg/kg oral dog: LDLo: 7500 mg/kg oral monkey: LDLo: 7000 mg/kg rabbit skin: LD50: 20 g/kg inhalation monkey: LCLo: 1000 ppm inhalation rat: LC50: 64,000 ppm/4H (Sax et al., 1989) Metabolism and methanol toxicity vary markedly between species (Jacobsen and Martin, 1986). In primates and humans, methanol causes metabolic acidosis and eye damage. However, in non-primate laboratory animals it acts as a depressant of the CENTRAL National SN (Gosselin et al., 1984; Rowe, 1982). 7.2.3 Relevant in vitro Data Several enzyme systems involved in the metabolism of methanol have been reported, including specific formaldehyde dehydrogenases found in numerous species and tissues (Tephly and 1984). The rate of oxidation of washing out is regulated by the hepatic concentration of tetrahydrofolate (Eells et al., 1982) 1982) Section 6.4). This observation may explain some differences in methanol toxicity between species and justifies therapeutic trials of folic acid (Jacobsen and McMartin, 1986). Formate can inhibit the activity of cytochrome C oxidase in pristine mitochondria, in subsenochondrial particles and in isolated cytochrome aa3. Inhibition increases with the reduction of pH (Nicholls, 1976). This finding may explain the formation of lactate, which occurs at the late stage of severe methanol poisoning (Jacobsen and McMartin, 1986). 7.2.4 TLV Workplace Standards (limit threshold): 200 ppm (Sax et al., 1989) TWA (weighted average) OSHA:200 ppm 260 mg/m3 NIOSH: 800 ppm/15 minutes short-term exposure limit of 7.2.5 Acceptable daily intake of food additives and contaminants The Committee recommended that methanol should be used as a solvent in food only for extraction purposes, and recommended a maximum concentration of 8 ppm in food. There are no data. Methanol is associated with birth defects in rats after both oral (Infurna et al., 1986) and inhalation exposure (Nelson, et al., 1985; Rogers et al., 1993; Bologna et al., 1994). There are no data. The main metabolic interaction occurs with ethanol, and is detailed in sections 6.4 and 10.6. 8.1.1 Sampling and sampling 8.1.1.1.1 Collection of blood and urine samples must be collected to identify methanol and folic acid. 8.1.2 Storage of laboratory samples and samples 8.1.2.1 Toxicological analyses of methanol and formate are stable in biological samples. Methanol, however, is a volatile substance and samples must be stored in tightly restricted containers. During all procedures, precautions should be taken to minimize the loss of alcohol by evaporation. If long-term storage is required prior to analysis, it is recommended to keep samples frozen at -20 degrees Celsius 8.2.2 Test for biological samples 8.2.2.1 Simple quality tests Fast procedures for detecting methanol in the blood (point test): no protein serum filter (1 ml) is added to the 0.1 ml KMnO4 solution (5 g in 100 ml H2O). The test tube should be gently tilted and, after 5 minutes, enough powdered sodium bisulfite should be added to decolorize the permanganate. Freshly prepared solution of chromotropic acid (0.2 ml, 0.5 g at 100 ml H2O) and 6 ml concentrated H2SO4 should be added and mixed and heated in a boiling water bath for 5 minutes. The red-purple color is positive and specific to methanol. 8.2.2.3 Simple quantitative quantification methods can be done with a coloratheria of 570 nm. This method involves measuring the intensity of color after methanol oxidation by formaldehyde, and then developing color by reacting formaldehyde with chromotropic acid (CTA). Availability of ethanol oxidized methanol procedure followed by the development of the CTA color if the calibration curve was established using pure methanol methanol Thus, this method can be used for initial detection of methanol, but is unsuitable for monitoring blood methanol levels during anti-dose ethanol treatment (false results) (Tietz, 1986). 8.2.2.4 Advanced quantitative methods of measuring methanol in the blood, gaschmatography is the method of choice. Plasma is diluted: equal volume of internal standard (2 ml methylethylketon in 1:1 water, procedure A (Baselt, 1982b) or: 1:10 with internal standard (3 ml 1 propanol in 1:1 water), procedure B (Blanke, 1975). It is highly recommended to use a pre-column. Column 125 KK; detector 200 degrees Celsius, nitrogen flow rate, 17 ml/minute. Procedure: introduction of 0.5 ml sample; storage time: methanol 0.6 minutes; (ethanol 0.9 minutes); 2.3 minutes. Procedure B: gas chromatographer with interchangeable glass injector, sleeve and flame ionization detector; 5' x 1/4 glass column containing 10% polyethylene glycol (PEG) 400 per 100 to 120 Anaknom 80. Injector 110 degrees Celsius, column 85 degrees Celsius, 125 C detector, nitrogen flow rate, 70 ml/minute Procedure: injecting 1 ml sample; Storage time: methanol 0.585 minutes, ethanol 0.654 minutes, internal standard 6 minutes. A quality control sample containing 1 g/l of each chemical is analyzed daily. 8.2.2.5 Other specialized methods of measuring formate in blood and urine are available by several methods. The Enzyme Method Shaller and Tribeg (1985), modified by zuppi and Motalbetti (1986), is accurate (cv by 1 mmol / 1.1.5%), simple, This should be the method of choice for large-scale screening in the event of a methanol poisoning epidemic (Soppi and Montalbetti, 1986) if the gas chromatographic definition of methanol cannot be met. In addition, it is easy to give out the enzyme definition of a comrade (zuppi and Montalbetti, 1986). : Formic acid oxidizes NED in the presence of dehydrogenase (FDH). The amount produced by NADH, measured at 340 nm, is equivalent to the amount acids are present in the sample. HCOOH and NAD+ FDH CO2 - NADH - H+ Optimal PH for FDH is between between and 7.5; the reaction occurs at ambient temperature. Formate can be measured in any biological fluid (urine, plasma, serum, or whole blood) deproteinized with perchloric acid 0.33 mmol/L Apparatus: Hitachi 705 (Boheringer Mannheim; Rotochem CFA 2000 (Kontron s.p.a.). Detailed procedure and its advantages over other enzyme methods are discussed by zuppi and Montalbetti (1986). HPLC principle: Phamic acid is separated on the column (250 mm x 4 mm) nucleosyl - 10 C18, having as a mobile phase a dual distilled H2O with pH, set at 3.53 sulfuric acid (flow 1 ml/minute pressure on the column 75 bar). Eluate is controlled at 210 Nm. The retention time at pH 3.53 for famic acid is 3.03 minutes. Gas chromatography of gas chromatography of pulpic acid can be performed by two procedures: degradation of formaldehyde with benzoic acid or: oxidation of fodder acid to CO and catalytic reduction on the pole to methane; methane is measured by ionizing the flame. 8.2.3 Interpretation of toxicological tests in late stages (severe metabolic acidosis) all ingested methanols can be metabolized and therefore cannot be detected in serum (Jacobsen, 1984). Methanol: To convert conventional units (mg/L) into mmol/L, divide methanol concentrations by 32.04 (Osterloh et al., 1986). Normal blood concentrations: Normal blood methanol concentrations derived from endogenous production and dietary sources, about 15 mg/L (0.05 mmol/L) (range from 2 to 30 mg/L) (Baselt, 1982a). Toxic concentrations in the blood: Recorded concentrations of plasma methanol zgt:5000 mg/L (157 mmol/L) (Gosselin et al., 1984; Gordfoot, 1982). Deadly concentrations: Concentrations of methanol in the blood are not necessarily a reliable predictive index (Baselt, 1982a; Jacobsen and McMartin, 1986). Toxicity clearly correlates with the degree of metabolic acidosis, i.e. the accumulation of pulpic acid. Bennet et al. (1953) reported blood concentrations of 0 to 3900 mg/L (0 to 122 mmol/L) (average 1,300 mg/L or 40 mmol/L) in 11 surviving patients, and concentrations from 0 to 4000 mg/L (0 to 125 mmol/L) (average 1600 mg/L or 50 mmol/L) in 7 who died during treatment (Bennet et al., 1953). Formic acid: Convert conventional units (mg/L) to milliequivalents in L divides formate concentrations 45.02 (Osterloh et al., 1986). Normal blood concentrations: From the literature and experience of collective poisoning of methanol-falsified wine in Italy in 1986, the concentration of pulp acid in normal subjects can vary from 0 to 18 mg/L. According to Baselt (1982a), the concentration of 1982a blood formic acid in normal subjects averages about 5 mg/L, and Osterich et (1986) report that normal formate concentrations are considered to be less than 12 mg/L. In workers exposed to production, the level of mimic acid in the blood increased from 3.2 to 3.2 in pre-received samples up to 7.9 mg/l by evening (Baselt, 1982a). Toxic concentrations in the blood: It has recently been suggested that serum concentrations are more correlated with clinical condition than methanol concentrations, and that the concentrations of the format are a more direct indicator of toxicity (Osterloh et al., 1986; Bozza-Marrubini et al., 1987; Jacobsen et al., 1983a,b; McMartin et al., 1980). It can be expected that blood concentrations exceeding 200 mg/L will lead to eye injuries or

